

SYNTHESIS OF DITERPENE ANALOGUES OF CARDENOLIDES

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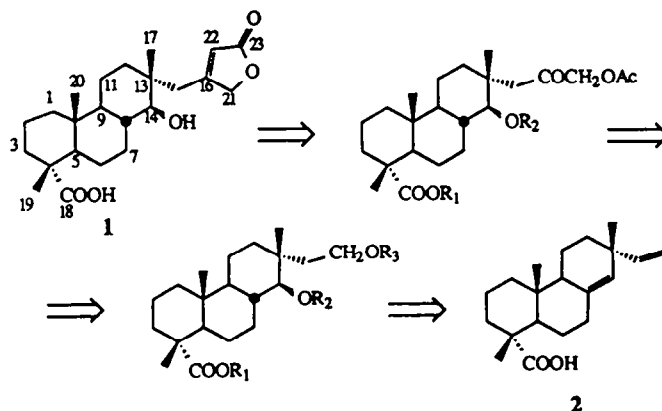
Abstract: The synthesis of diterpene analogues of cardenolides from sandaracopimaric acid has been achieved. Funtionalization in C₁₄ and C₁₆ was carried out by hydroboration-oxidation and the formation of the butenolide ring was conducted through a Reformatsky-type reaction.

The search for new inotropic analogues of cardenolides receives justification in the need for obtaining substances with a better therapeutic index and in establishing the structural requirements for a positive interaction with the inotropic receptor of cardiac glycosides^{1,2,3}.

With these aims in mind, we have developed a line of research directed towards obtaining compounds that maintain, on different structural residues, a butenolide and a hydroxyl group in an arrangement similar to that of the cardiac genins⁴. This situation can be achieved on a diterpene skeleton such as isopimarane, which shows a close structural relationship with the steroid system.

In the present work we describe the synthesis of the lactone **1** from sandaracopimaric acid **2**, a compound that can be obtained from several species of the families Cupresaceae and Pinaceae⁵.

In the retrosynthetic study (scheme I), we planned the construction of the butenolide ring from an acetoxymethylketone grouping, which can be obtained from a carboxyl group on C₁₆. The introduction of this group and that of the hydroxyl group at position C₁₄ can be carried out by successive functionalization of positions 16 and 14, through selective hydroboration-oxidation of sandaracopimaric acid, in the sequence already described by us⁶.

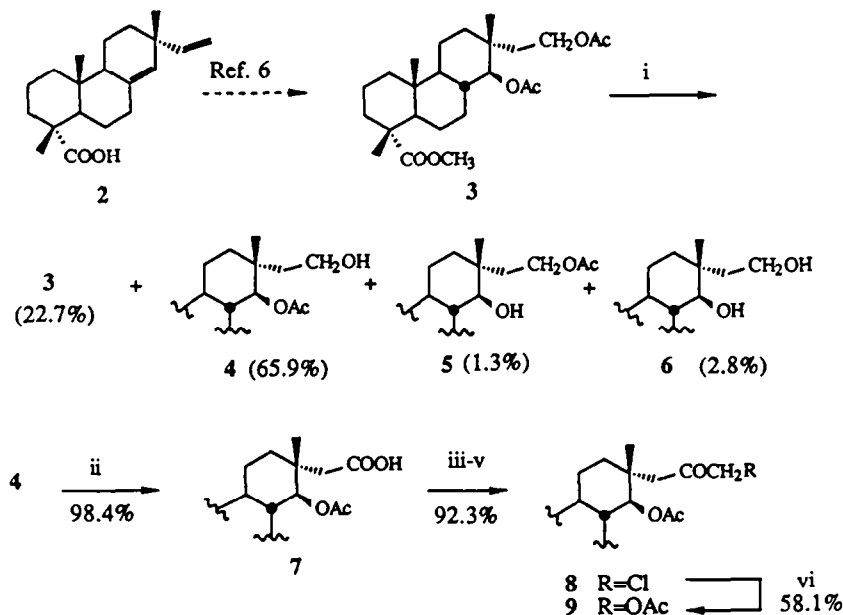


Scheme I

RESULTS AND DISCUSSION

The synthesis of compound **1** was carried out as depicted in schemes II, III and IV, starting with the conversion of sandaracopimaric acid (**2**) into the ester diacetate **3**⁶.

Deacetylation of **3** at C₁₆ was carried out by selective saponification, taking into account the easier accessibility of the acetate on C₁₆ with respect to the acetate on C₁₄. Thus, by treatment with NaHCO₃-H₂O/MeOH (1:3) over 40 h at room temperature, a mixture of products was obtained with methyl 14 β -acetoxymethyl-16-hydroxy-13-*epi*-pimarane-18-oate (**4**) as the expected major component. Using other proportions of NaHCO₃-H₂O/MeOH (1:5 or 1:2) and using longer reaction times a smaller yield of **4** was obtained. The minor products **5** and **6**, once acetylated and mixed with **3**, were subjected several times to treatment with base leading to a 93% total yield in compound **4** (scheme II).



i. NaHCO₃-H₂O/MeOH (1:3); ii. Jones; iii. SOCl₂; iv. CH₂N₂; v. HCl(g); vi. NaOAc/Ac₂O

Scheme II

The oxidation of **4** with Jones reagent (CrO₃/H₂SO₄) led to the acetoxycarboxylic acid **7**, from which the side chain was lengthened by one carbon atom. The transformation of **7** into its acid chloride followed by treatment with diazomethane afforded the derived diazoketone, which was transformed into the chloromethylketone **8** by reaction with HCl (g)⁷. Substitution of the chloride by acetate led to the acetoxymethylketone **9**⁸ (scheme II).

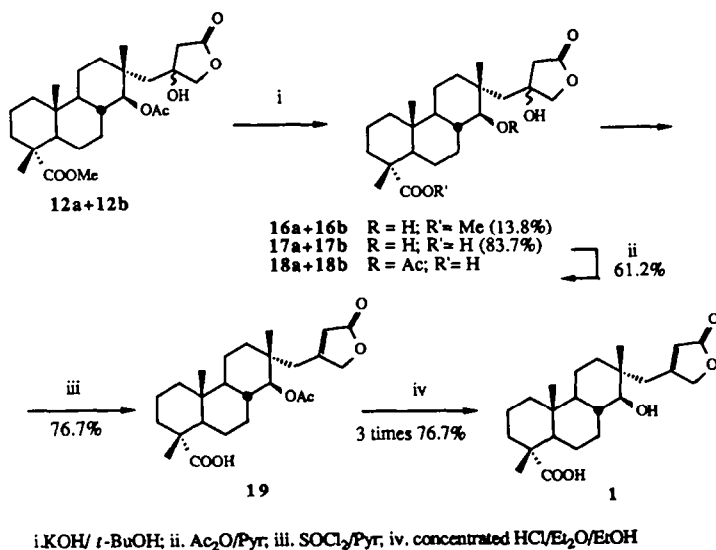
Scheme III

Selective hydrolysis of the C₁₄-acetate of **11** in acid medium¹² was produced without affecting the carbomethoxy group at C₁₈ nor the butenolide, to give a mixture of **14** and **11**. After chromatographic separation, the recovered portion of **11** was subjected to several hydrolysis-separation processes, producing an almost quantitative transformation into **14**. The best yields were obtained with ether/ethanol/concentrated hydrochloric acid at a proportion of 1:1:0.6. Substances **12a+12b** and **13a+13b** are mixtures of epimers at C₁₆ that differ in some signals of their NMR spectra (Tables III and IV).

When the product of the Reformatsky reaction was not subjected to any basic treatment, component separation proved to be more difficult, because apart from producing the above mentioned **10**, **11** and **12a+12b**, compounds **15a+15b** were also present in the reaction product. Saponification of this mixture yielded **12a+12b** and under the same conditions **13a+13b** was converted into the butenolide **11**, which accounts for the low percentage isolated of **13a+13b** and the absence of **15a+15b** in the Reformatsky reaction described above. Another reaction carried out to increase the butenolide yield was the dehydration of **12a+12b** with SOCl_2 .

The attempts to carry out the hydrolysis of the methyl ester on C_{18} in the presence of the butenolide on C_{15} were first made with $\text{HOAc}/\text{quinoline}$, because the hydrolysis of axial and equatorial esters has been described under these conditions¹³ and in our hands it had been effective on methyl sandaracopimarate, and secondly with $\text{KOH}/t\text{-BuOH}$. In both cases, the butenolide was degraded in a similar fashion as the cardiac glycosides¹⁴.

As a consequence, the synthesis of **1** was performed from **12a+12b**. Treatment of **12a+12b** with saturated $\text{KOH}(t\text{-BuOH})$ led to mixtures of epimers **16a+16b** and **17a+17b**. The mixture of acetylated epimers **18a+18b** was dehydrated to **19**, whose hydrolysis in acid medium occurred in a similar way to that described for **11**, yielding a mixture of the final product **1** and of **19** (scheme IV).



Scheme IV

The structure of compounds **1-19** was established by means of their spectroscopic properties¹⁵. Compounds **1** and **14** are currently being studied regarding their inotropic activity and the results will be published elsewhere.

TABLE I. ¹H NMR data (200MHz) for compounds 1, 3-11, 14 and 19.

H	1*	3	4	5	6	7	8	9	10	11	14	19
14	2.97 d (9.5)	4.50 d (10.1)	4.50 d (10.1)	2.94 d (9.7)	2.95 d (9.8)	4.56 d (10.2)	4.56 d (10.2)	4.53 d (9.7)	3.51 d (11.1)	4.52 d (10.0)	2.88 d (9.7)	4.53 d (10.0)
15	2.69 d (13.2)					2.21 d (13.7)	2.40 s	2.23 s	2.27 s	2.30 d (13.8)	2.53 d (13.5)	2.31 d (13.8)
15	2.49 d (13.2)					2.13 d (13.7)				2.23 d (13.8)	2.41 d (13.5)	2.22 d (13.8)
16		4.09 t (7.6)	3.67 m	4.19 t (7.4)	3.69 m							
17	1.06 s	0.98 s	0.93 s	0.91 s	0.88 s	1.07 s	1.07 s	1.07 s	0.98 s	1.00 s	0.94 s	1.00 s
19	1.39 s	1.16 s	1.15 s	1.19 s	1.17 s	1.16 s	1.17 s	1.16 s	1.18 s	1.16 s	1.17 s	1.16 s
20	0.90 s	0.95 s	0.88 s	0.90 s	0.88 s	0.98 s	0.89 s	0.88 s	0.95 s	0.89 s	0.88 s	0.90 s
21	4.89 s									4.72 m (1.9)	4.77 d	4.73 m
22	6.06 s									5.90brs	5.86brs	5.91brs
OMe		3.62 s	3.62 s	3.66 s	3.64 s	3.63 s	3.62 s	3.62 s	3.65 s	3.63 s	3.65 s	
OAc		2.06 s 2.01 s	2.06 s	2.05 s		2.07 s	2.06 s	2.06 s		2.08 s		2.08 s
							4.03 s	2.14 s				
								4.55 s				

* Pyr
CDCl₃, δ (ppm), from internal TMS. J Hz.

TABLE II. ¹³C NMR data (50.3 MHz) for compounds 1, 3-11, 14 and 19.

C	1*	3	4	5	6	7	8	9	10	11	14	19
1	37.8	38.2	38.2	38.2	38.2	38.1	38.2	38.1	38.0	38.2	38.2	38.2
2	18.8	18.1	18.1	18.1	18.1	18.1	18.0	18.0	18.1	18.1	18.1	18.0
3	35.9	36.8	36.7	36.9	36.2	36.7	36.4	36.7	37.0	36.8	36.9	36.9
4	47.6	47.5	47.5	47.5	47.4	47.5	47.4	47.4	47.6	47.5	47.6	47.2
5	49.9	49.3	49.2	49.6	49.6	49.2	49.1	49.1	51.0	49.2	49.5	49.0
6	24.5	23.6	23.8	23.9	24.0	23.6	23.5	23.5	23.4	23.6	23.8	23.8
7	31.8	30.3	30.4	31.0	31.1	30.2	30.2	30.2	30.6	30.3	30.8	30.4
8	39.3	36.5	36.6	38.2	38.0	36.7	36.1	36.4	34.2	37.0	38.8	37.1
9	54.7	54.2	54.2	54.4	54.5	54.0	53.9	54.0	54.5	54.0	54.2	54.1
10	36.5	36.2	36.2	35.6	36.9	36.2	36.6	36.1	36.6	36.2	36.3	36.2
11	19.9	19.2	19.2	19.3	19.4	19.2	19.0	19.0	19.6	19.2	19.5	19.2
12	38.7	35.0	35.0	36.2	38.2	35.0	34.7	34.6	33.6	35.5	35.8	35.5
13	40.0	36.8	36.9	37.3	37.7	37.6	38.0	38.1	40.9	38.6	39.4	38.6
14	80.4	82.6	82.8	81.5	81.5	82.6	82.4	82.7	91.5	82.4	81.1	82.6
15	40.6	39.3	43.5	40.2	46.5	45.5	49.1	48.7	46.2	40.1	40.4	40.1
16	169.3	60.8	58.6	61.4	58.3	176.8	200.6	202.2	200.8	166.4	167.8	166.5
17	17.9	17.5	18.1	16.9	15.8	17.0	17.2	17.0	17.5	18.1	17.5	18.0
18	181.4	178.9	179.0	179.2	179.4	179.0	178.8	178.8	179.1	178.9	179.2	184.5
19	17.5	16.7	16.7	16.7	16.7	16.7	16.6	16.6	16.6	16.7	16.7	16.5
20	14.8	14.4	14.4	14.5	14.5	14.1	14.3	14.3	14.7	14.5	14.6	14.5
21	75.4									74.8	75.1	74.8
22	116.3									118.9	118.4	118.9
23	174.3									173.6	174.0	173.8
OMe		51.7	51.7	51.7	51.7	51.6	51.7	51.7	51.9	51.8	51.8	
OAc		20.8(2)	20.8	21.0		20.8	20.7	20.7		20.3		20.5
		170.8(2)	171.7	170.9		171.0	170.9	170.9		170.4		171.0
Others							49.6		20.3			
								69.1				
								169.9				

* Pyr
CDCl₃, δ (ppm), from internal TMS.

TABLE III. ^1H NMR data (200MHz) for the pairs of epimeric compounds 12-13, 15-18.

H	12a	12b	13a	13b	15a	15b	16a	16b	17a	17b*	18a	18b
14	4.89 d (9.9)	4.92 d (9.9)	4.78 d (10.9)	4.69 d (10.9)	4.58 d (10.0)	4.61 d (10.0)	3.12 d (9.8)		3.19 d (9.6)	4.89 d (10.0)	4.91 d (9.8)	
17	0.95 s		1.00 s	1.01 s	1.04 s		1.00 s	0.95 s	1.20 s	1.16 s	0.96 s	
19	1.16 s		1.16 s	1.17 s	1.16 s		1.18 s		1.38 s		1.16 s	
20	0.89 s		0.88 s	0.95 s	0.89 s		0.88 s		0.89 s	0.85 s	0.90 s	
21	4.10 d (9.8)	3.90 d (9.7)	4.10 d (10.4)	4.18 d (10.6)	4.00 d (11.3)	3.99 d (11.3)	4.03 d (9.7)	4.23 d (11.5)	4.22 d (9.1)	4.42 d (9.1)	4.12 d (9.6)	3.95 d (10.2)
21	4.44 d (9.8)	4.06 d (9.7)	4.45 d (10.4)	4.40 d (10.6)	4.16 d (11.3)	4.08 d (11.3)	4.25 d (9.7)	4.38 d (11.5)	4.50 d (9.1)	4.62 d (9.1)	4.46 d (9.6)	4.06 d (10.2)
22	2.48 s (17.2)	2.44 d (17.2)	2.68 d (17.9)	2.10 d (18.0)	2.55 s (17.9)	2.60 s (18.0)	2.65 s (17.9)	2.45 d (17.9)	2.97 s (17.9)	2.85 s (17.9)	2.47 d (17.9)	2.48 d (17.9)
22	2.48 s (17.2)	2.75 d (17.2)	3.00 d (17.9)	3.10 d (18.0)			2.65 s (17.9)	2.60 d (17.9)			2.50 d (17.9)	2.68 d (17.9)
OMe	3.62 s		3.62 s		3.62 s		3.65 s					
OAc	2.12 s		2.03 s	2.07 s	2.07 s						2.08 s	
Others					1.28t (7.1)							
					4.16 c (7.1)	4.15 c (7.1)						

* Pyr
 CDCl_3 , δ (ppm), from internal TMS. J Hz.

TABLE IV. ^{13}C NMR data (50.3 MHz) for the pairs of epimeric compounds 12-13, 15-18.

C	12a	12b	13a	13b	15a	15b	16a	16b	17a	17b*	18a	18b
1	38.1		38.2		38.2		38.2		38.6		38.0	
2	18.0		18.1		18.1		18.2		18.8		18.0	
3	36.7		36.8		36.6		37.0		37.6		36.9	
4	47.5		47.5		47.6		47.6		47.7		47.2	
5	49.2		49.3		49.3		49.4		49.9		49.0	
6	23.5		23.6		23.7		23.8		24.6		23.6	
7	30.5		30.4		30.6		30.7		31.9		30.6	
8	36.5		36.8		36.6		38.5		39.0		36.6	
9	54.0		53.9		54.2		54.2		54.8		53.9	
10	36.2		36.4		35.8		36.3		36.5		36.2	
11	19.1		19.2		19.3		19.1		19.7		19.2	
12	34.4		36.2		36.2		38.2		39.3		38.0	
13	38.9		38.8		38.9		39.2		40.1		38.9	
14	83.2		83.5		84.0		82.4		82.2	82.1	83.2	
15	46.3	45.1	41.3		43.5	42.9	52.8	53.8	52.1	52.9	46.3	45.3
16	77.1	76.8	83.3		73.3		75.7	76.4	76.5	77.1	76.4	77.0
17	20.2		17.6		18.8		16.7		17.6		21.0	
18	179.0		179.0		179.0		179.3		181.4		184.3	
19	16.6		16.8		16.8		16.7		17.6		16.4	
20	14.4		14.5		14.4		14.6		14.8		14.4	
21	80.2	81.0	77.1	75.0	69.5	70.1	81.1	79.9	81.5	80.7	80.2	80.6
22	45.7	43.9	44.3	45.0	45.7	45.1	43.8	45.0	44.7	45.5	45.8	44.3
23	175.1		174.3		179.0		175.2		177.1		175.2	
OMe	51.8		51.9		51.8		51.9					
OAc	21.0		20.7	22.1	20.8	20.9					20.4	
	172.9		170.4	171.1	170.4	171.4					173.2	
Others					14.1	60.6						

* Pyr
 CDCl_3 , δ (ppm), from internal TMS.

EXPERIMENTAL

General: The solvents and reagents were purified and dried by standard techniques. Mps are uncorrected. The IR spectra were taken in 4% CHCl_3 solution and NMR spectra were obtained in CDCl_3 solution (200 MHz for ^1H , 50.3 MHz for ^{13}C), unless otherwise stated. Chemical shifts are reported in ppm (δ) downfield from internal TMS. Optical rotations were measured at 20°C on a digital polarimeter. Mass spectra were obtained under electron impact (70 eV) and ultraviolet spectra were obtained in methanol.

Methyl 14 β -acetoxy-16-hydroxy-13-*epi*-pimaran-18-oate (4):

70 ml of saturated NaHCO_3 (aq) were added to a stirred solution of **3** (7.4 g, 17 mmol) in 210 ml of methanol and maintained with vigorous stirring for 30 h at 25°C. The mixture was then diluted with water, extracted with EtOAc and washed with brine. After the usual work up the crude product (7.1 g) was chromatographed over silica gel to yield: unreacted **3** (1.7 g, 22.7%; Hexane/EtOAc 6:4); **5** (75 mg, 1.3%; Hexane/EtOAc 6:4); **4** (3.7 g, 65.9%; Hexane/EtOAc 6:4) and **6** (170 mg, 2.8%; EtOAc).

Acetylation of **5** and **6** gave a further amount of diacetate **3**, that was again saponified. After seven times, the partial saponification of **3** yielded 6.2 g of **4** (93%).

4. $[\alpha]^{20}_D$ (λ): +8.7° (589), +9.3° (578), +10.5° (546), +19.3° (436), +32.8° (365), $c = 1.14\%$ (CHCl_3).

IR: 3520, 1740, 1270, 1160, 1040 cm^{-1} .

^1H -NMR: Table I. ^{13}C -NMR: Table II.

Methyl 16-acetoxy-14 β -hydroxy-13-*epi*-pimaran-18-oate (5):

$[\alpha]^{20}_D$ (λ): -4.0° (589), -4.3° (578), -4.6° (546), -7.0° (436), -9.3° (365), $c = 0.93\%$ (CHCl_3).

IR: 3500, 1745, 1730, 1250, 1140, 1060 cm^{-1} .

^1H -NMR: Table I. ^{13}C -NMR: Table II.

Methyl 14 β ,16-dihydroxy-13-*epi*-pimaran-18-oate (6):

Mp = 136-137°C. $[\alpha]^{20}_D$ (λ): -16.2° (589), -17.4° (578), -20.2° (546), -31.0° (436), -47.4° (365), $c = 0.98\%$ (CHCl_3).

IR: 3220, 1720, 1230, 1140, 1100, 1060 cm^{-1} .

^1H -NMR: Table I. ^{13}C -NMR: Table II.

14 β -acetoxy-18-methoxy-18-oxo-13-*epi*-pimaran-16-oic acid (7):

A stirred solution of alcohol **4** (6 g, 16.2 mmol) in acetone (160 ml) at 0°C was titrated with Jones reagent ($\text{CrO}_3/\text{H}_2\text{SO}_4$). The mixture was extracted with EtOAc when the orange colour persisted. After the usual work up 6.1 g of **7** (98.4%) were obtained.

7. Mp = 176-178°C. $[\alpha]^{20}_D$ (λ): +19.4° (589), +20.6° (578), +23.2° (546), +41.2° (436), +67.3° (365), $c = 1.03\%$ (CHCl_3).

IR: 3300-2500, 1740, 1720, 1250, 1040 cm^{-1} .

^1H -NMR: Table I. ^{13}C -NMR: Table II.

Methyl 16-acetoxy-14 β -hydroxy-16-oxo-13-*epi*-pimaran-18-oate (9):

Acetoxyacid **7** (6 g, 14.7 mmol) was converted into the acylchloride with thionylchloride (25 ml) in dry benzene. An ethereal solution of the acylchloride was added dropwise to an excess of ethereal diazomethane and maintained for 3 h between -5°C and 0°C. Afterwards, a HCl(g) current was bubbled through the reaction mixture for 1 h. By usual work up **8** (6 g, 92.3%) was obtained.

5.9 g (13.4 mmol) of **8**, acetic anhydride (119 ml) and anhydrous sodium acetate (20 g, 217 mmol) were refluxed under N_2 for 4 h. After extraction, the organic layer was evaporated and the residue was chromatographed over SiO_2 to give **9** (3.6 g, 58.1%).

Methyl 14 β -acetoxy-16-chloromethyl-16-oxo-13-*epi*-pimaran-18-oate (8):

Mp = 142–144°C. $[\alpha]^{20}_D(\lambda)$: +22.0° (589), +23.5° (578), +27.3° (546), +53.6° (436), +112.4° (365), $c = 1.13\%$ (CHCl₃).

IR: 1740, 1730, 1250, 1040, 980, 910, 870 cm⁻¹.

¹H-NMR: Table I. ¹³C-NMR: Table II.

9. Mp = 82–86°C. $[\alpha]^{20}_D(\lambda)$: +10.2° (589), +10.9° (578), +12.5° (546), +23.1° (436), +43.1° (365), $c = 0.98\%$ (CHCl₃).

IR: 1745, 1720, 1250, 1170, 1130, 1030 cm⁻¹.

¹H-NMR: Table I. ¹³C-NMR: Table II.

Reformatsky reaction

A- A solution of ethyl bromoacetate (96 ml, 0.86 mol) and α -acetoxyketone 9 (450 mg, 0.96 mmol) in dry benzene was added dropwise, to a stirred mixture of active granulated zinc (450 mg, 6.8 mmol) in dry benzene. The mixture was refluxed for 6 h, and the zinc compound was decomposed as usual with dilute HCl (aq). Extraction with EtOAc yielded 450 mg of reaction product. After several saponifications, acetylations and chromatographies, 10 (9.8%), 11 (17.8%), 12a+12b (22.8%), and 15a+15b (5.6%) were obtained.

18-methoxy-18-oxo-13-*epi*-pimaran-14 β ,16-olide (10):

Mp = 191–193°C. $[\alpha]^{20}_D(\lambda)$: -61.3° (589), -64.5° (578), -73.5° (546), -126.3° (436), $c = 1.04\%$ (CHCl₃).

IR: 1780, 1720, 1250, 1170, 1070, 1045, 990 cm⁻¹.

¹H-NMR: Table I. ¹³C-NMR: Table II.

Methyl 14 β -acetoxy-16-carboxymethyliden-16-hydroxymethyl-13-*epi*-pimaran-18-oate γ -lactone (11):

$[\alpha]^{20}_D(\lambda)$: +24.9° (589), +26.3° (578), +29.9° (546), +52.4° (436), $c = 1.22\%$ (CHCl₃).

IR: 1790, 1755, 1725, 1640, 1250, 1180, 1040 cm⁻¹.

UV λ_{\max} nm (ϵ): 212 (20300).

¹H-NMR: Table I. ¹³C-NMR: Table II.

Methyl 14 β -acetoxy-16-carboxymethyl-16-hydroxy-16-hydroxymethyl-13-*epi*-pimaran-18-oate γ -lactone (12a+12b):

IR: 3380, 1780, 1720, 1250, 1140, 1100, 1050, 1020, 820 cm⁻¹.

¹H-NMR: Table III. ¹³C-NMR: Table IV.

Methyl 14 β -acetoxy-16-acetoxymethyl-16-ethoxycarbonylmethyl-16-hydroxy-13-*epi*-pimaran-18-oate (15a+15b):

IR: 3480, 1740, 1720, 1250, 1100, 1030, 970 cm⁻¹.

¹H-NMR: Table III. ¹³C-NMR: Table IV.

B- The reaction was carried out in the conditions described in A with zinc (2.6 g, 39.7 mmol), 9 (2.6 g, 5.6 mmol) and ethyl bromoacetate (35 ml, 314 mmol). The mixture was refluxed under N₂ for 6 h and the zinc compound was decomposed as usual with dilute HCl (aq). Extraction with EtOAc yielded 3 g of reaction product. After column chromatography 10 (100 mg, 5.2%) and another complex mixture (2.2 g) were obtained. This latter mixture was stirred in NaHCO₃-H₂O/MeOH (100/120 ml) for 72 h at 25°C and was extracted with EtOAc to give 1.25 g of reaction product. The aqueous layer was acidified and after extraction yielded 960 mg.

By acetylation and chromatography of both fractions: 11 (430 mg, Hexane/EtOAc 65:35) and 12 (280 mg, Hexane/EtOAc 1:1) were isolated from the neutral fraction and 13 (38 mg, Hexane/EtOAc 6:4) and 12 (590 mg, Hexane/EtOAc 1:1) were obtained from the acid fraction.

Methyl 14 β ,16-diacetoxy-16-carboxymethyl-16-hydroxymethyl-13-*epi*-pimaran-18-oate γ -lactone (13a+13b)

IR: 1785, 1745, 1725, 1250, 1040, 980 cm^{-1} .

$^1\text{H-NMR}$: Table III. $^{13}\text{C-NMR}$: Table IV.

Dehydration of 12a+12b:

A solution of 12a+12b (45 mg, 0.097 mmol), pyr (1.2 ml) and SOCl_2 (0.5 ml) was stirred under N_2 for 45 min at 25°C. The solution was diluted with water, extracted with EtOAc and after the usual work up, the crude product was chromatographed (hexane/EtOAc 65:35) to give 11 (32 mg, 74%).

Methyl 16-carboxymethyliden-14 β -hydroxy-16-hydroxymethyl-13-*epi*-pimaran-18-oate γ -lactone (14):

11 (85 mg, 0.19 mmol) in Et_2O /EtOH/concentrated HCl (1:1:0.6) was maintained for 4 days at room temperature and then extracted with EtOAc. By chromatography (hexane/EtOAc 6:4) of the reaction product, unreacted 11 (37.2 mg, 43.4%) and 14 (31.8 mg, 40.8%) were obtained.

14. $[\alpha]^{20}_{\text{D}}$: +8.6° (589), +10.9° (578), +19.1° (546), $c = 0.70\%$ (CHCl_3).

IR: 3600, 1790, 1720, 1640, 1260, 1110, 1040, 990, 900, 870 cm^{-1} .

MS: 404 (M^+ , 21), 345 (61), 307 (61), 247 (99), 229 (90), 173 (17).

UV λ_{max} nm (ϵ): 217 (5050).

$^1\text{H-NMR}$: Table I. $^{13}\text{C-NMR}$: Table II.

16-carboxymethyliden-14 β -hydroxy-16-hydroxymethyl-13-*epi*-pimaran-18-oic γ -lactone acid (1):

Saponification of methylester group in C_{18} : 12 (250 mg, 0.56 mmol) in saturated KOH(*t*-BuOH) (20 ml) was stirred under N_2 at 100°C for 1 h and at 40°C for 12 h. The reaction mixture was extracted with EtOAc to give 16a+16b (31.4 mg, 13.8%) and the aqueous layer was acidified with 2N HCl (aq) and extracted with EtOAc yielding 17a+17b (184 mg, 83.7%).

Methyl 16-carboxymethyl-16-hydroxymethyl-14 β ,16-dihydroxy-13-*epi*-pimaran-18-oate γ -lactone (16a+16b):

IR: 3600, 3350, 1785, 1720, 1260, 1110, 1050, 1040, 1030, 990 cm^{-1} .

$^1\text{H-NMR}$: Table III. $^{13}\text{C-NMR}$: Table IV.

16-carboxymethyl-16-hydroxymethyl-14 β ,16-dihydroxy-13-*epi*-pimaran-18-oic acid γ -lactone (17a+17b):

IR (1% KBr): 3600-2400, 3250, 1785, 1735, 1700, 1100, 1050, 990 cm^{-1} .

$^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}$). Table III. $^{13}\text{C-NMR}$: Table IV.

By acetylation of 17a+17b (145 mg) and chromatography over deactivated SiO_2 (5% H_2O), 18a+18b (98 mg, 61.2%) were isolated.

18a+18b (95 mg, 0.19 mmol), pyr (2.4 ml) and SOCl_2 (1 ml) were stirred under N_2 , in the same conditions of dehydration of 12a+12b, to give 19 (70 mg, 76.7%).

Hydrolysis of 19: The reaction of 19 (65 mg, 0.15 mmol) under the same conditions employed in the acidic hydrolysis of 11, yielded unreacted 19 (32 mg, 50%) and 1 (22 mg, 38.1%).

16-carboxymethyliden-14 β -hydroxy-16-hydroxymethyl-13-*epi*-pimaran-18-oic acid γ -lactone (1):

$[\alpha]^{20}_{\text{D}}$: -3.4° (589), -3.7° (578), -4.1° (546), $c = 1.00\%$ (Pyr).

IR (1% KBr): 3600-2700, 3500, 1725, 1695, 1630, 1200, 1100, 1030, 990 cm^{-1} .

MS: 390 (M^+ , 2), 345 (5), 275 (10), 293 (10), 247 (31), 229 (21), 173 (8).

$^1\text{H-NMR}$ ($\text{C}_5\text{D}_5\text{N}$). Table III. $^{13}\text{C-NMR}$: Table IV.

14 β -acetoxy-16-carboxymethyl-16-hydroxy-16-hydroxymethyl-13-*epi*-pimaran-18-oic acid γ -lactone (18a+18b):

IR : 3600-2400, 3420, 1780, 1735, 1700, 1260, 1140, 1125, 1060, 1030, 970 cm^{-1} .

$^1\text{H-NMR}$: Table III. $^{13}\text{C-NMR}$: Table VI.

14 β -acetoxy-16-carboxymethyliden-16-hydroxymethyl-13-*epi*-pimaran-18-oic acid γ -lactone (19):

IR: 3500-2400, 3500, 1795, 1750, 1730, 1700, 1650, 1200, 1100 cm^{-1} .

$[\alpha]^{20}_{\text{D}}$ (λ): +25.3° (589), +26.2° (578), +30.4° (546), +53.5° (436), $c = 0.97\%$ (CHCl_3).

UV λ_{max} nm (ϵ): 213 (10617).

$^1\text{H-NMR}$: Table I. $^{13}\text{C-NMR}$: Table II.

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REFERENCES

1. Thomas, R.E.; *Burger's Medicinal Chemistry* 4th Ed. Part III, Ed. Wolf, M.E., Jonh Wiley & Sons, New York 1981, p. 45.
2. Fullerton, D.S.; *Wilson and Gisvold's Textbook of Organic Medicinal and Pharmaceutical Chemistry*, Ed. Doerge, R.F., 8^o Ed, J.B. Lippincott CO, Philadelphia 1982, p. 712.
3. Fullerton, D.S.; Ahmed, K.; From, A.H.L.; Mc Parland, R.H. and Griffin, J.F. *Topics in Molecular Pharmacology*, Vol. 3. Ed. Burgen, A.S.V.; Roberts, C.G.K. and Tute, M.S. Elsevier, Amsterdam 1986, p. 261.
4. San Feliciano, A.; Medarde, M.; Caballero, E.; Hebrero, B.; Tomé F.; Montero, M.J. and Prieto, P. *Eur. J. Med. Chem.* 1990, 25, 0000.
5. Hegnauer, R. *Chemotaxonomie der Pflanzen*, Vol. 1. Birkhauser, Basel. 1962, p. 367.
6. San Feliciano, A.; Medarde, M.; Tomé F.; Caballero, E.; Hebrero, B.; Miguel del Corral, J.M. and Barrero, A.F. *Tetrahedron* 1989, 45, 1815.
7. Walker, J. *J. Chem. Soc.* 1940, 1304.
8. Schmitt, J.; Suquet, M.; Cornoy, P.; Boitard, J.; Callet, J.; Clim, T. and Le Meur, J. *Bull. Soc. Chim.* 1986, 953.
9. Wadsworth, W.S, JR and Emmons, W.D. *J. Am. Chem. Soc.* 1968, 83, 1733.
10. Fried, J.; Linville, R.G. and Elderfield, R.C. *J. Org. Chem.* 1942, 7, 362.
11. Nef, J.U. *Liebigs Ann. Chem.* 1914, 403, 204.
12. Kreiser, W.; Warnwcke, H.U. and Neef, G. *Liebigs Ann. Chem.* 1973, 2071.
13. Aranda, G. and Fetizon, M.; *Synthesis* 1975, 330.
14. Paist, W.D.; Blout, E.R.; Uhle, F.C. and Elderfield, R.C. *J. Org. Chem.* 1941, 6, 273.
15. San Feliciano A.; Medarde, M.; Tomé, F.; Hebrero, B. and Caballero, E. *Magn. Reson. Chem.* 1989, 27, 1116.